

# Pathogenic and Molecular Characterization of *Fusarium oxysporum* f.sp. *ciceri* Causing Chickpea Wilt through ISSR Markers

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**Abstract**—In the present investigation the pathogenic and genetic variability was assayed, amongst the seven isolates of *Fusarium oxysporum* f.sp. *ciceri* (Foc) collected from different agro-climatic zones of Maharashtra State, India. The isolates of *Fusarium oxysporum* f.sp.*ciceri* were confirmed by SCAR marker which yielded 1.5 KD band. The pathogenicity of each isolate was confirmed using the wilt susceptible chickpea genotype JG-62. On the basis of pathogenic ability the isolates were grouped as highly pathogenic (FOC-2, FOC-5, FOC-6), strongly pathogenic (FOC-1, FOC-3) and moderately pathogenic (FOC-4, FOC-7). Eight Inter Simple Sequence Repeats primers (ISSRs) were used to determine the genetic variability in seven isolates *Fusarium oxysporum* f.sp. *ciceri*. The seven primers produced 80 scorable bands. Of 80 bands, 73 bands were polymorphic and average level of polymorphism was 91.25 per cent. In UPGMA analysis, Foc-1 (Wardha) was found to have higher value of similarity coefficient (0.8375) whereas Foc-2 (Lonar) was found to have lower value of similarity coefficient (0.4625). The isolates of *Fusarium oxysporum* f.sp.*ciceri* were grouped into two major clusters. First group, cluster-A includes isolates belonging to Wardha, Washim and Lonar. Second group, cluster-B includes Nashik, Ahmednagar, Rahuri and Pune. It shows that Foc-1 (Wardha) have higher value of similarity coefficient with Foc-3 (Washim) whereas Foc-2 (Lonar) have lower value of similarity coefficient with Foc-5 (Ahmednagar). The similarity matrix indicated that seven isolates of *Fusarium oxysporum* f.sp.*ciceri* exhibited in between 46-84 per cent similarity coefficient.

**Keywords**—*Fusarium oxysporum* f.sp. *ciceri*, ISSR Markers, Pathogenic variation.

## I. INTRODUCTION

Chickpea (*Cicer arietinum* L.) is one of the most important pulse crop grown worldwide. It is reported to be infested by

more than 52 pathogen (Nene et al., 1984). The losses due to *Fusarium oxysporum* f. sp. *ciceri* had been reported to the tune of 10 to 100% Grewal et al. (1974). Being soil borne in nature use of host plants resistance is the most practical and economical approach for management of *Fusarium* wilt of chickpea (Haware, 1990). The pathogen is monophyletic in nature, complete one life cycle in a year, but the occurrence of races in the form of variability had been demonstrated by Haware and Nene (1982). The studies the pathogenic variability revealed that the chickpea wilt pathogen *Fusarium oxysporum* f.sp. *ciceri* had four distinct physiological races in India (Haware and Nene, 1982). Hence, it becomes necessary to ascertain the molecular variability amongst the isolates of *Fusarium oxysporum* f.sp. *ciceri* in order to have the specific breeding programme, it is necessary to determine the genetic variability among the isolates of *Fusarium oxysporum* f.sp.*ciceri* at molecular level, so as the breeding programme for race specific wilt resistant varieties could be initiated. The present investigation will helps to understand the genetic variability among the isolates of *Fusarium oxysporum* f.sp. *ciceri* in Maharashtra, which in turn will be useful for its exploitation for regional race specific breeding programme for developing wilt resistance variety. The various workers had demonstrated that molecular markers viz. RAPD (Bayraktar and Dollar 2007) ISSR (Akram et.al.2014, Borrego-Benjumea et al. 2014) ITS-RFLP (Arpita Das et.al.2012, Gayatri Gurjar et al. 2009), AFLP (Mamta Sharma et al. 2009) RAPD and ISSR (Madhuri Katkar et.al 2015) were useful in characterizing genetic variation in a wide range of organisms. In the present investigation ISSR molecular markers were used to analyze the genetic variability of *Fusarium oxysporum* f.sp. *ciceri* isolates collected from different agro-climatic zones of Maharashtra, India.

## II. MATERIALS AND METHODS

### Fungal isolates

A total seven isolates of *F. oxysporum* f. sp. *ciceris* (FOC) were isolated from wilt-infected chickpea plants from different agro-climatic zones of Maharashtra (Table-1) were designated as FOC-1 (Wardha), FOC-2 (Lonar), FOC-3 (Washim), FOC-4 (Nashik), FOC-5 (Ahmednagar), FOC-6 (Rahuri) and FOC-7 (Pune). Isolations were made from the wilted plants on potato dextrose agar (PDA) medium. The fungus was identified according to the identification keys of *F. oxysporum* f.sp.*ciceri*.

### Molecular Characterization of *F. oxysporum* f.sp.*ciceri*.

The *Fusarium oxysporum* f.sp. *ciceri* specific Sequence Characterized Amplified Region (SCAR) primer pair Foc0-12 f-GGCGTTTCGCAGCCTTACAATGAAG and Foc0-12 r-GACTCCTTTTCCCGAGGTAGGTCAGAT were used for the confirmation of the isolates (Jimenez Gasco and Jimenez Diaz, 2003)

### Pathogenicity test

The pathogenicity test of isolates of *F. oxysporum* f.sp. *ciceri* isolated from the different agro-climatic zones of the Maharashtra state, India; were tested by using susceptible cultivar JG-62. The seven isolates of *F. oxysporum* f.sp. *ciceri* were purified by hyphal tip method were mass multiplied separately on sand maize meal medium. The soil for experiment taken was autoclaved for 3 day consequently at 30 lbs pressure for 30 minutes. The mass multiplied inoculum was mixed with the soil in 1:10 proportion and filled in the presterilized pots. Five seeds of susceptible chickpea cultivar JG-62 were sown in pots each isolate in 3 replications. The per cent emergence and post-emergence mortality at 30 DAS was recorded and total mortality was calculated. The seedlings maintained in sterilized soil without inoculums were served as control. The pathogenicity test was confirmed by proving Koch's Postulate and pathogens were confirmed as *Fusarium oxysporum* f.sp *ciceri*.

### Grouping of *Fusarium oxysporum* f. sp. *ciceri* isolates

The isolates of *F. oxysporum* f. sp. *ciceri* were tentatively divided in five groups based on their pathogenic ability on cv. JG-62 given as per Paul et.al. (2001).

Category	Per cent wilt/root rots
1. Nonpathogenic (NPI)	0%
2. Weakly pathogenic (WPI)	1-20%
3. Moderately pathogenic (MPI)	21-50%
4. Strongly pathogenic (SPI)	51-70%
5. Highly pathogenic (HPI)	> 70%

### Genomic DNA extraction

The pure culture of seven isolates of *Fusarium oxysporum* f.sp *ciceri* were grown on Potato Dextrose Broth (200 g peeled potato, 20 g dextrose, 1000 ml distilled water) for 7 days ( $27 \pm ^\circ\text{C}$ ). The mycelial mat for each isolates was harvested. The DNA was extracted following CTAB method of DNA extraction by Murray and Thompson (1980) with little modifications. The DNA obtained after extraction was confirmed by running it on 0.8% agarose gel containing ethidium bromide at 0.5 mg/ml in a horizontal gel electrophoresis system. 2  $\mu\text{l}$  of genomic DNA of each isolate + 3 $\mu\text{l}$  loading dye + 5 $\mu\text{l}$  sterile water loaded in each well. After completion of 5 cm run, the gel was observed under UV light and the DNA yield and quality was confirmed

### ISSR Analysis

The eight ISSR primers supplied by "BIOSERVE" were used to amplify the DNA. The DNA amplification was carried out in a thermal cycler (Eppendorf, Germany) with the conditions as described by Kelly et al. (1994) as follows: one cycle of initial denaturation at  $94^\circ\text{C}$  for 10 minutes, followed by 35 cycles of denaturation at  $94^\circ\text{C}$  for 45 sec, annealing at  $50-54^\circ\text{C}$  (as per primers) for 45 sec and primer extension at  $72^\circ\text{C}$  for 2 minutes. A final extension at  $72^\circ\text{C}$  for 10 minutes was carried out at the end of the cycles. Amplifications was performed in a total volume of 25  $\mu\text{l}$  of reaction, containing 10 mM of Tris-HCl ( $\text{pH}$  8.8), 50 mM of KCl, 2.5 mM of  $\text{MgCl}_2$ , 0.24  $\mu\text{M}$  of primer, 0.2mM of dNTPs, and 1 U of Taq polymerase. The eight ISSR primers with di- and tri- nucleotide repeats were analyzed. The annealing temperatures were adjusted as per G+C content of the primers.

### Electrophoresis of ISSR-PCR product:

The ISSR analysis was carried out in vertical gel electrophoresis. The PCR products were separated eletrophoretically in 1.5 percent agarose gel. The ISSR-PCR product (7 $\mu\text{l}$ ) with 3  $\mu\text{l}$  loading dye in each well was loaded along with 1 kb DNA ladder (5  $\mu\text{l}$ ) with 3  $\mu\text{l}$  loading dye in the first well. The gel was run for at 80V.

### Gel documentation and Data analysis:

The gel images were captured and visualizaed in gel documentation system (Biorad). The data was scored as the presence (1) or absence (0) of individual band for each isolates in PCR-ISSR analysis of each isolate of *Fusarium oxysporum* f.sp. *ciceri*. The data was used to generate similarity coefficient using simple matching coefficient based on ISSR bands pattern. The Dice coefficient between each pair of accessions were then used to construct a

dendrogram using the Unweighted Pair Group Method with Arithmetic Mean Average (UPGMA).

### III. RESULTS AND DISCUSSION

#### Molecular characterization of isolates of *Fusarium oxysporum* f.sp. *ciceri*:

All the isolates of the races yielded the single 1.5 kb band (Plate-2 ) with the SCAR marker, hence confirms the culture of *Fusarium oxysporum* f.sp. *ciceri* were as per the result of Jimenez Gasco and Jimenez- Diaz (2003).

#### Pathogenicity and Grouping of isolates

The results (Table -2 and Table-3) indicates that all the among seven isolates of *Fusarium oxysporum* f. sp. *ciceri*, all proved to be pathogenic to susceptible cultivar JG-62. Isolate from FOC-2 was highly virulent (100%) and caused wilting and death of the seedlings in 20-22 days. Whereas remaining isolates viz. FOC-1 (66.66%), FOC-3 (60.00%), FOC-4 (50.00%), FOC-5 (92.85%), FOC-6 (84.61%) and FOC-7 (42.00%) respectively. On the basis of the reactions the seven isolates *F. oxysporum* f.sp. *ciceri* FOC-4, FOC-7 has been classified as Moderately Pathogenic (21-50%), FOC-1, FOC-3 as Strongly Pathogenic (51-70%) and FOC-2, FOC-5, FOC-6 as Highly Pathogenic (> 70%). On the basis of pathogenic ability by Paulkar et al. (2002) reported the virulence of four isolates of *F. oxysporum* f. sp. *ciceri* under laboratory conditions. Isolate Foc-3 showed 100 % mortality on JG-62 (highly pathogenic), isolates Foc-1 and Foc-4 showed 70 and 90 per cent mortality respectively. Whereas, Foc-2 isolate showed moderately pathogenic reaction with 60 % mortality. Barhate et al. (2006) reported per cent mortality among these seven isolates three isolates were highly pathogenic (71-100%), two strongly pathogenic and two were moderately pathogenic.

#### Molecular Characterization

The genetic diversity (Table-4 Plate-1 and 3) was studied amongst the seven isolates of *Fusarium oxysporum* f.sp. *ciceri* using eight ISSR primers. Among them seven primers produced 80 scorable bands. Out of 80 bands, 73 bands were polymorphic and average level of polymorphism was 91.25 per cent. The primer (GA)<sub>9</sub>RY amplified maximum 15 bands within the size 408bp-13984bp, while primer (AG)<sub>8</sub>T and (AG)<sub>8</sub>YT amplified minimum of 9 bands within the size 550bp to 13628bp and 394bp to 2766bp respectively. The six ISSR primers (AG)<sub>8</sub>T (13628bp); (ATG)<sub>6</sub> (7706bp); (AG)<sub>8</sub>G (712bp); (AG)<sub>8</sub>YT (587bp) (CA)<sub>8</sub>RT (2675bp) and (AC)<sub>8</sub>YT (1384bp), (1210bp) showed 7 monomorphic bands.

#### Binary similarity matrix for ISSR analysis

On the basis of calculated similarity matrix, the similarity between two genotypes was predicted. The genotypes showing similarity index of “1” are presumed to be 100 per cent similar while that of “0” are 100 per cent genetically dissimilar. In the dendrogram (Table 1 & 5 , Fig-1) the similarity matrix indicated that seven isolates of *Fusarium oxysporum* f.sp. *ciceri* exhibited in between 46-84 per cent similarity coefficient. It was clear that in cluster A, the three isolates FOC-1 (Wardha) and FOC-2 (Washim) were grouped which showed the 83.75% similarity. The isolates FOC -1 (Wardha) and FOC -2 (Washim) falls under the agro-ecological zone viz. Moderate rainfall zone VIII and assured rainfall zone which has the soil PH ranging from 7 to 7.5 and the soil type Vertisols and Inceptisols. Whereas the isolate FOC -3 (Lonar) showed 46.25% similarity with FOC -1 (Wardha) and FOC -2 (Washim) falls in cluster A and the variation may be due to high alkaline pH of soil which had sustained the meteorite impact in the past. In cluster-B, it includes isolates FOC -4 (Nashik), FOC -5 (Ahmednagar), FOC -6 (Rahuri, Ahmednagar) and FOC -7 (Pune). From the dendrogram it is cleared that two isolates FOC -4 (Nashik), FOC -5 (Ahmednagar) exhibits 66% similarity level, both the agro-ecological zones are adjacent to each other with moderately alkaline soils with pH ranging from 7.4 to 8.4. The FOC -6 (Rahuri, Ahmednagar) isolates showed the 61 % similarities with the rest of isolates, falls in the scarcity zone, where as FOC -7 (Pune) falls in the Western Maharashtra Plain Zone which as the lateritic to black soil has pH 6 to 7 which are poor in the potash and phosphorus has the 58% similarities with rest of all the isolates. Borrego-Benjumea et al. (2014) studied genetic variation in several soil borne *Fusarium* spp. caused *Fusarium* crown and root rot of asparagus. Ninety-three *Fusarium* isolates, obtained from plant and soil samples collected from commercial asparagus fields in southwestern Ontario, were identified as *Fusarium oxysporum* (65.5%), *F. proliferatum* (18.3%), *F. solani* (6.4%), *F. acuminatum* (6.4%), and *F. redolens* (3.2%) based on morphological or cultural characteristics and polymerase chain reaction (PCR) analysis with species-specific primers. The inter simple-sequence repeat (ISSR) PCR analysis of the field isolates revealed considerable variability among the isolates belonging to different *Fusarium* spp. Akram et al. (2014) studied the genetic variation amongst ten different isolates of *Fusarium oxysporum* f. sp. *lycopersici* causing wilt in tomato. These isolates were collected from infected tomato plants from different tomato fields. Genetic finger printing of all *F. oxysporum* f. sp. *lycopersici* isolates was performed by using ISSR markers. Dendrogram based on the ISSR

analysis divided all *F. oxysporum* f. sp. *lycopersici* isolates in two major groups.

Thus it clearly indicates that the genesis of soil, pH, presence of types of nutrients and elements in the soil plays important roles in evolution of the soil borne plant pathogen their variation, multiplication adoption and survival that had been clear from the present investigation.

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Table.1: Collection of isolates from different agro-climatic zones of Maharashtra State, India

Sr.	Agro climatic zone	District of Collection	Designated as	Soil Type
1.	Moderate rainfall zone (zone-VIII)	Wardha	FOC-1	Black soil derived from basalt rock, Medium to heavy in texture alkaline to reaction. Low lazing areas are rich, fertile.
2.	Assured rainfall zone (zone-VII)	Lonar, Dist. Buldhana	FOC-2	Soil colour ranges from black to red type Vertisols, Entisols, Inceptisols PH 7-7.5
3.	Assured rainfall zone (zone-VII)	Washim	FOC-3	
4.	Transition zone-II	Nashik	FOC-4	Soil grayish black, Moderately alkaline 7.4 to 8.4 lowest layer is murum strata Fair in NPK content, well drained good for irrigation
5.	Scarcity zone	Ahmednagar	FOC-5	Vertisols with Montmorillonite clay, poor in nitrogen low to medium in phosphate and well supplied in potash
6.	Scarcity zone	Ahmednagar (Rahuri)	FOC-6	
7.	Western Maharashtra Plain zone	Pune	FOC-7	Soils are reddish brown to black tending to lateritic Ph-6 to 7, well supplied in Nitrogen and low in phosphorus and potash.

Table.2 : Pathogenicity reaction of isolates of *Fusarium oxysporum* f.sp.ciceri

Isolates	No. of plants	Germination	% Germination	Wilted plants	% Wilting	Wilting (DAI)
FOC-1	15	12	80.00	08	66.66	24
FOC-2	15	15	100.00	15	100.00	22
FOC-3	15	10	66.66	06	60.00	25
FOC-4	15	14	93.33	7	50	25
FOC-5	15	14	93.33	13	92.85	26
FOC-6	15	13	86.66	11	84.61	25
FOC-7	15	14	93.33	6	42.86	22
Control	15	15	100.00	00	00	00

Table .3: Grouping of *F. oxysporum* f.sp. ciceri isolates on the basis of pathogenicity .

Sr.	Group (% Mortality)	<i>F. oxysporum</i> f.sp.ciceri
1	Non-Pathogenic (0%)	(0)
2	Weakly Pathogenic (1-20%)	(0)
3	Moderately Pathogenic (21-50%)	FOC-4,FOC-7
4	Strongly Pathogenic (51-70%)	FOC-1,FOC-3
5	Highly Pathogenic (> 70%)	FOC-2,FOC-5,FOC-6

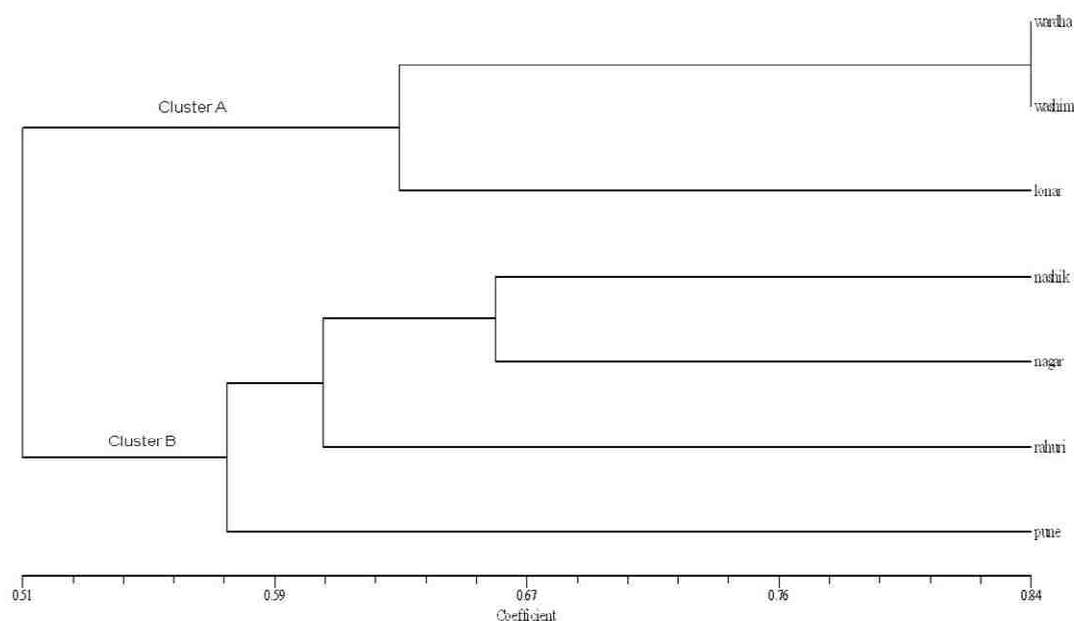


Table.4: The ISSR primers and their polymorphism

Sr. No.	Primer	Total Bands	Polymorphic bands	% Polymorphism
1	(GA) <sub>9</sub> RY	15	15	100
2	(AG) <sub>8</sub> T	9	8	88.88
3	(ATG) <sub>6</sub>	10	9	90
4	(AG) <sub>8</sub> G	12	11	91.66
5	(AG) <sub>8</sub> YT	9	8	88.88
6	(CA) <sub>8</sub> RT	11	10	90.90
7	(AC) <sub>8</sub> YT	14	12	85.71
Total		80	73	91.25

Table.5: Similarity coefficient for ISSR analysis

Foc-1 Wardha	Foc-2 Lonar	Foc-3 Washim	Foc-4 Nashik	Foc-5 Ahmednagar	Foc-6 Rahuri	Foc-7 Pune
1.0000						
0.5875	1.0000					
0.8375	0.6750	1.0000				
0.5375	0.5000	0.6000	1.0000			
0.4750	0.4625	0.4875	0.6625	1.0000		
0.4625	0.5500	0.4750	0.6000	0.6125	1.0000	
0.5000	0.5125	0.5375	0.6375	0.5250	0.5625	1.0000

Fig.1: The ISSR UPGMA dendrogram of seven isolates of *F. oxysporum* f. sp. *ciceri* based on Jaccard's Similarity Coefficient

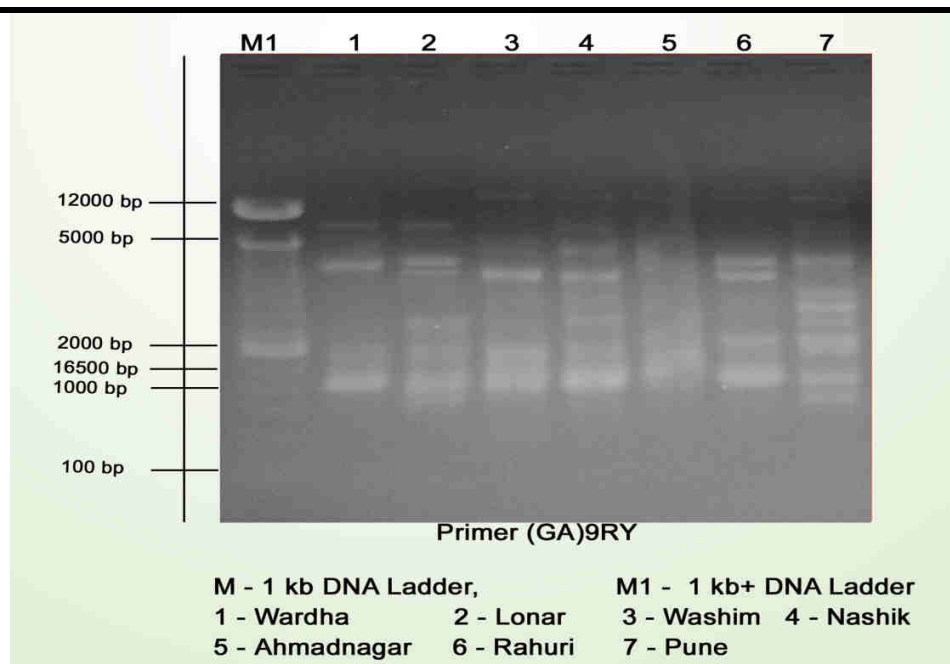
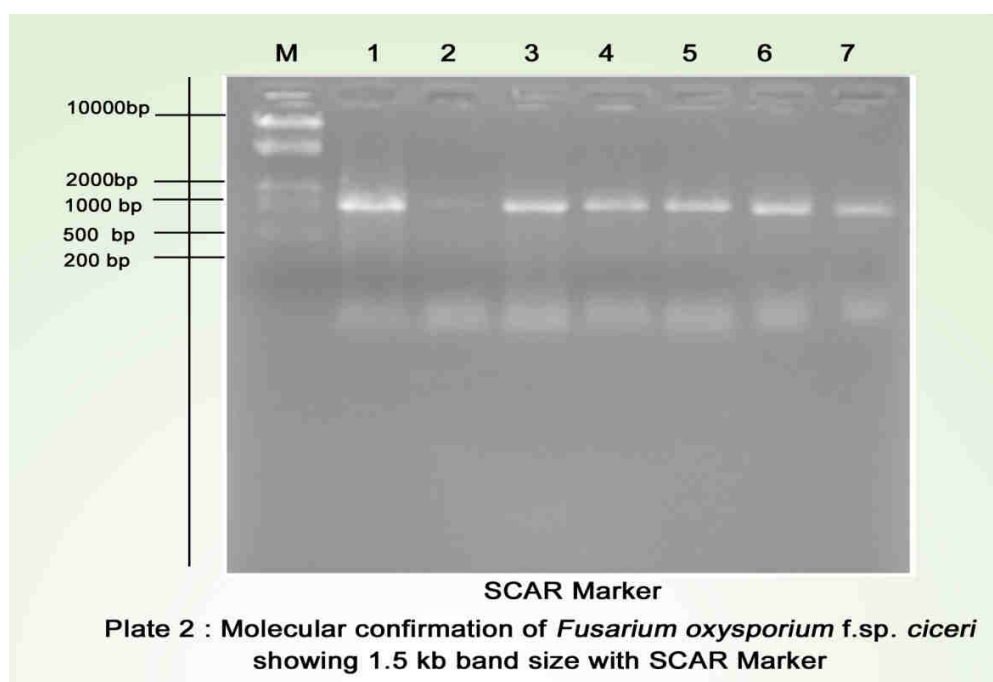


Plate 1: ISSR banding pattern of Primer (GA)<sub>9</sub>RY



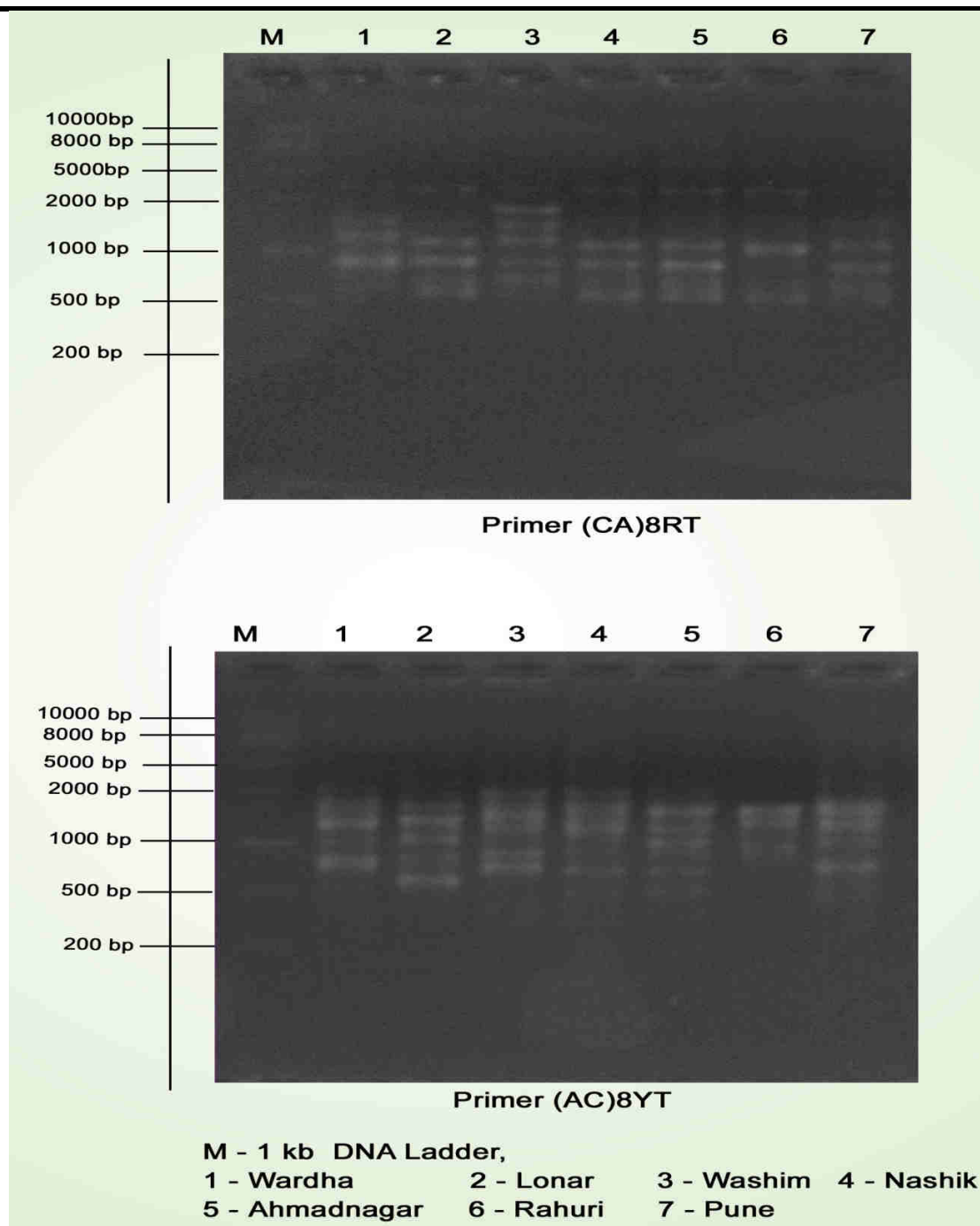


Plate 3: ISSR banding pattern of Primer (CA)<sub>8</sub>RY and (AC)<sub>8</sub>YT